Vol. 88, No. 3, 1979 BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS
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ERRATA

Volume 87, Number 3, April 13, 1979

In "Endogenous, Cyclic 3'-5' AMP-Dependent Phosphorylation of Human Red Cell Pyruvate Kinase," by Joëlle Marie, Lydie Tichonicky, Jean-Claude Dreyfus, and Axel Kahn, pp. 862-868, through a printer's error, page 867 was omitted. For the convenience of our readers, the correct pages 867 and 868 are reproduced on the following pages.

subunits of liver enzyme. Moreover the identical dephosphorylation induced by subtilisin on pyruvate kinases from red cells incubated with or without cAMP argues against the hypothesis that two distinct sites of phosphorylation could exist, sensitive to cAMP-dependent or cAMP independent protein kinases. Trypsin is by far less active in dephosphorylating phosphorylated pyruvate kinase than subtilisin, although the sequence of the phosphorylated site includes an arginyl residue (14). This finding seems to indicate that the main cleavage point attacked by trypsin is not located in the same position as that attacked by subtilisin. Trypsin would mimic the physiological maturation of the phosphorylatable L'_4 precursor into the phosphorylatable L_4 enzyme found in the liver (7, 8). In contrast, subtilisin would be especially active in removing the phosphorylatable site of both L' and L subunits. Though endogenous cAMP-dependent phosphorylation of erythrocyte membrane proteins is known for several years (15), it is the first time that endogenous cAMP-dependent phosphorylation of a red cell enzyme is documented. The functional significance of such a phenomenon in cells devoid of gluconeogenic pathway and with hormone-insensitive adenylcyclase (16) remains obscure and further studies are needed to clarify this problem.

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ERRATA (continued)

Volume 86, Number 4, February 28, 1979

In "Distribution of Hexokinase Isoenzymes Depending on a Carbon Source in Saccharomyces cerevisiae," by Haruhiro Muratsubaki and Takurou Katsume, pp. 1030-1036, the following should be added with respect to ethanol-grown cells: All ethanol-grown cells were cultivated on the medium containing both 3% ethanol and 0.5% glucose as carbon source.

Volume 87, Number 1, March 15, 1979

In "Stimulation of the Conversion of Penicillin N to Cephalosporin by Ascorbic Acid, α-Ketoglutarate, and Ferrous Ions in Cell-Free Extracts of Strains of Cephalosporium acremonium," by D. J. Hook, L. T. Chang, R. P. Elander, and R. B. Morin, pp. 258-265, on page 258, in the abstract, the concentrations of ascorbic acid and ferrous ions should read 3.8 mM and 0.075 mM, respectively, instead of 3.8 μM and 0.075 μM. Page 259, paragraph 2, line 6, should read "The suspension was incubated" rather than "inoculated." Page 263, Table 6, the concentrations given for $FeSO_4$ should read "100 mmoles" instead of "100 μ moles." Page 265, line 2, "isopenicillin N" should read "penicillin N."

ERRATA (continued)

Volume 87, Number 2, March 30, 1979

In "Binding versus Biological Activity of Clostridium perfringens Enterotoxin in Vero Cells," by James L. McDonel and Bruce A. McClane, pp. 497-504, on page 499, line 6 from the bottom, "1.29 x 10^6" should read "0.78 x 10^6." On page 500, line 4, "1.35 x 10^5" should read "0.81 x 10^5."

Volume 87, Number 2, March 30, 1979

In "Metal Ion Induced Conformational Changes in Concanavalin A: Evidence for Saccharide Binding to One Metal Free Structure" by C. A. Stark and A. D. Sherry, pp. 598-604 , we reported that single additions of Co^{2+} or Zn^{2+} to apoconcanavalin A induces the structural conversion of this protein to the structural form which binds with saccharides. These erroneous data apparently resulted from a Ca^{2+} contamination in our protein stock. A reexamination of these experiments with carefully purified protein and buffer solutions shows that Co^{2+} or Zn^{2+} alone will not induce the conformational transition. However, less than stoichiometric amounts of Ca^{2+}, i.e., Ca^{2+}/Con A ratio of 0.3 to 0.6, purposely added with excess Co^{2+} produces the fully active saccharide binding structure. Thus, Ca^{2+} apparently plays a catalytic role in the conformational transition. The Co^{2+} ion in this form is not labile as evidenced by little change in the visible absorption spectrum upon addition of EDTA and the Zn^{2+}-Con A and Co^{2+}-Con A derivatives bind with α-methylunbelliferyl-D-mannopyranoside with an affinity of $3.3\pm0.4 \times 10^4 M^{-1}$ at 25°C. These derivatives containing a metal ion only in S1 are metastable and unfold to apo structure with the reported rate constants. Occupancy of S2 by Ca^{2+} is therefore not required for saccharide binding but is required to stabilize this structural form of the protein.